

## **An** *N***-Aryl-Substituted Oxazolidinone-Containing Ketone-Catalyzed Asymmetric Epoxidation with Hydrogen Peroxide as the Primary Oxidant**

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Asymmetric epoxidation of various olefins with an *N*-arylsubstituted oxazolidinone-containing ketone as catalyst and hydrogen peroxide as the primary oxidant has been investigated, and up to 96% ee was obtained.

Dioxiranes have proven to be valuable agents for the epoxidation of olefins.<sup>1,2</sup> Typically they are prepared by using a ketone and Oxone (potassium peroxymonosulfate). In our earlier studies on epoxidation with fructose-derived ketone **1**, we have shown that  $H_2O_2$  coupled with a nitrile activator presents a viable alternative to Oxone for the formation of dioxiranes and subsequent epoxidation of olefins (Scheme 1).<sup>3</sup> High yields and ee values were obtained for a wide variety of trans- and trisubstituted olefins. Further studies have shown that some other ketones can be effective for the epoxidation with the  $RCN-H<sub>2</sub>O<sub>2</sub>$  system.<sup>4,5</sup> In this epoxidation, peroxyimidic acid is likely the active oxidant for the formation of the dioxirane (Scheme 1).<sup>6</sup> Hydrogen peroxide  $(H_2O_2)$  is a highly desirable oxidant because of its high active oxygen content and its

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reduction product being water. $7-9$  In addition, ketone-catalyzed epoxidation reactions with  $H_2O_2$  require less solvent and salts than those with Oxone and do not require slow addition of oxidant.

Fructose-derived ketone **1** has been shown to be an effective catalyst for the epoxidation of a wide variety of trans- and trisubstituted olefins.2b,c,10 During our further studies, we have found that oxazolidinone-containing ketones **2** gave high ee values for substrates such as *cis*-olefins,<sup>11,12</sup> styrenes,<sup>11,13</sup> and certain trisubstituted olefins11,14 which are not effective with ketone 1. The question is whether the  $RCN-H<sub>2</sub>O<sub>2</sub>$  system can also be extended to epoxidation with ketone **2**. Herein, we wish to report our efforts on this subject.

Early on in the study it became apparent that ketone **2a** would not be useful under these conditions because of its rapid

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<sup>(7)</sup> For a general reference on hydrogen peroxide see: Strukul, G. *Catalytic Oxidations with Hydrogen Peroxide as Oxidant*; Kluwer Academic Publishers: New York, 1992.

<sup>(8)</sup> For leading reviews on epoxidation of olefins with hydrogen peroxide see: (a) Grigoropoulou, G.; Clark, J. H.; Elings, J. A. *Green Chem.* **2003**, *5*, 1. (b) Noyori, R.; Aoki, M.; Sato, K. *Chem. Commun*. **2003**, 1977. (c) Burgess, K.; Lane, B. S. *Chem. Re*V. **<sup>2003</sup>**, *<sup>103</sup>*, 2457. (d) Kelly, D. R.; Roberts, S. M. *Biopolymers* **2006**, *84*, 74. (e) Matsumoto, K. *Yuki Gosei Kagaku Kyokaishi* **2006**, *64*, 869. (f) Arends, I. W. C. E. *Angew. Chem.*, *Int. Ed.* **2006***, 45, 6250.*



decomposition.15 Ketones **2b**,**c** are more readily available than **2a** and appeared to be more robust under the reaction conditions so further study was conducted with these catalysts. Initial studies were carried out with *cis*-*â*-methylstyrene as a test substrate. Under the optimum conditions for ketone **1**, epoxidation with ketone 2**b** using  $H_2O_2$  in CH<sub>3</sub>CN as solvent gave poor enantioselectivity (∼50% ee). Subsequently, various solvents were tested for the epoxidation with 5 mol % ketone **2b**. As shown in Table 1, *n-*BuOH was found to be among the best solvents (Table 1, entry 11). The ee was increased to 78% when the amount of CH<sub>3</sub>CN was reduced (Table 1, entry 12). After more optimization, it was found that 3.8 equiv of CH3- CN along with 3 equiv of  $H_2O_2$  in *n*-BuOH-0.30 M aq  $K_2CO_3$ in  $4 \times 10^{-4}$  M EDTA (1:1, v/v) gave the best results overall for a number of substrates.<sup>16</sup> In previous studies of various olefins, ketone **2c** often gave better overall results than **2b**, 13 so we decided to examine a variety of olefins with this catalyst (Table 2).17 In many cases the ee values obtained are within a few percent of if not as high as the corresponding ee values with Oxone as oxidant. Up to 96% ee was obtained (Table 2, entry 7). The drop in enantioselectivity observed in some cases (e.g. Table 2, entry 11) is likely due to a solvent effect since the asymmetric induction of ketone **2** is largely attributed to electronic and/or hydrophobic effects which are solvent dependent.<sup>11-14</sup>

During the course of study it was also observed that the ketones are susceptible to decomposition under the reaction conditions. The reaction pH with 0.30 M  $K_2CO_3$  is about 11.5, and at such a high pH  $\alpha$ -deprotonation of the ketone occurs subsequently eliminating acetone.<sup>18</sup> It is also possible that the





*<sup>a</sup>* The reactions were carried out with olefin (0.5 mmol), ketone **2b** (0.025 mmol), and 30%  $H_2O_2$  (0.20 mL, 2.0 mmol) in CH<sub>3</sub>CN (0.20 mL, 3.8 mmol)-solvent (0.75 mL)-0.6 M K<sub>2</sub>CO<sub>3</sub> in  $4 \times 10^{-4}$  M aqueous EDTA (0.75 mL) at  $0^{\circ}$ C (bath temperature). <sup>*b*</sup> The conversions and ee values were determined by chiral GC (Chiraldex B-DM). *<sup>c</sup>* CH3CN (0.10 mL, 1.9 mmol) used. *<sup>d</sup>* The reaction time is 4 h.

oxazolidinone is gradually hydrolyzed over time, presumably due to the nucleophilic action of peroxide anion.<sup>15</sup> The major consequence of this is that relatively unreactive olefins are difficult to drive to full conversion because the decomposition process(es) competes with the epoxidation process. However, for many substrates this system is a viable alternative to the Oxone protocol.

In summary, a method for asymmetric epoxidation with use of *N*-aryl oxazolidinone-containing ketones with  $H_2O_2$  as primary oxidant has been described. A variety of olefins can be epoxidized with good yields and ee values. Use of  $H_2O_2$ allows for use of less solvent, salts, and eliminates the need for slow addition of oxidant. The reactions are operationally simple, and in many cases give results similar to those obtained with Oxone.

## **Experimental Section**

The general experimental information is similar to that recently described.<sup>10b</sup> Hydrogen peroxide  $(H_2O_2)$  is potentially explosive, and although no incidents occurred in our experience, care must be taken in handling this compound. In the epoxidation reaction, EDTA is used to minimize the decomposition of  $H_2O_2$  catalyzed by any trace metals. Freshly purchased  $H_2O_2$  from Aldrich was used in this study. It was found that vigorous stirring is crucial for epoxidation efficiency, particularly for less reactive substrates. All epoxides are known except Table 2, entries 5 and 8, and gave satisfactory spectroscopic characterization.

**Representative Procedure for Asymmetric Epoxidation (Table 2, entry 2).** A mixture of 2-methyl-1-phenyl-1-propene (0.135 g, 1.00 mmol) and ketone **2c**<sup>21</sup> (0.051 g, 0.15 mmol) in *n*-BuOH (3.0 mL) was cooled to 0  $^{\circ}$ C with an ice bath. CH<sub>3</sub>CN (0.20 mL, 3.8) mmol) and 0.30 M K<sub>2</sub>CO<sub>3</sub> in  $4 \times 10^{-4}$  M aqueous EDTA (3.0) mL) were then added, followed by  $H<sub>2</sub>O<sub>2</sub>$  (30%, 0.30 mL, 3.0 mmol) with vigorous stirring at 0  $\degree$ C. The mixture was stirred at 0  $\degree$ C for

<sup>(15)</sup> For facile hydrolysis of oxazolidinone with LiOOH see: Evans, D. A.; Britton, T. C.; Ellman, J. A. *Tetrahedron Lett.* **1987**, *28*, 6141.

<sup>(16)</sup> Under these conditions, 28% conversion and 78% ee were obtained for *cis*-*â*-methylstyrene with ketone **2a**.

<sup>(17)</sup> It was observed that olefins containing hydroxy groups are usually poor substrates under the reaction conditions for reasons that are not well understood.

<sup>(18)</sup> The formation of acetone can be detected by NMR.

## [OC Note





*<sup>a</sup>* The reactions were carried out with olefin (1.0 mmol), ketone **2c** (0.10-0.30 mmol), CH3CN (0.20 mL, 3.8 mmol), *<sup>n</sup>*-BuOH (3.0 mL), aq 0.30 M K<sub>2</sub>CO<sub>3</sub> in 4  $\times$  10<sup>-4</sup> M EDTA (3.0 mL), and 30% H<sub>2</sub>O<sub>2</sub> (0.30 mL, 3.0 mmol) at 0 °C for the time indicated. <sup>*b*</sup> Absolute configuration was determined by comparing the measured optical rotations to the reported ones. *<sup>c</sup>* Conversion was determined by GC of the crude reaction mixture. *<sup>d</sup>* Enantioselectivity was determined by chiral GC (Chiraldex B-DM column). *<sup>e</sup>* Conversion was determined by 1H NMR of the crude reaction mixture. *<sup>f</sup>* Enantioselectivity was determined by chiral HPLC (Chiralcel OD column).

24 h and was then poured into petroleum ether and extracted with petroleum ether. The combined organic layers were washed with water and saturated aqueous  $Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>$ , dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, concentrated, and purified by flash chromatography (silica gel was buffered with  $1\%$  Et<sub>3</sub>N in petroleum ether; petroleum ether was used as eluent) to give 2,2-dimethyl-3-phenyloxirane as a colorless oil (0.121 g, 82% yield, 92% ee).

**Table 2, entry 1:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub>  $-40.0$  (*c* 0.77, CHCl<sub>3</sub>) (82% ee); 1H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.40-7.28 (m, 5H), 4.07

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(21) Ketones **2b** and **2c** used in this study were prepared by the PDC oxidation of the alcohol precursors (for less reactive olefins, best results were obtained with freshly recrystallized ketones). It was observed that lower conversions were obtained with the ketone prepared from the TEMPObleach oxidation of the alcohol.<sup>22</sup> It appears that some residual impurities from TEMPO-bleach oxidation somehow affect the epoxidation.

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(d,  $J = 4.4$  Hz, 1H), 3.39-3.32 (m, 1H), 1.10 (d,  $J = 5.2$  Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  135.7, 128.2, 127.7, 126.8, 57.8, 55.4, 12.7.

**Table 2, entry 2:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> +38.3 (*c* 0.85, C<sub>6</sub>H<sub>6</sub>) (92%) ee); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.40–7.28 (m, 5H), 3.87 (s, 1H), 1.49 (s, 3H), 1.08 (s, 3H); 13C NMR (100 MHz, CDCl3) *δ* 136.8, 128.2, 127.6, 126.6, 64.8, 61.3, 25.0, 18.2.

**Table 2, entry 3:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> +99.3 (*c* 0.46, EtOH) (88% ee); 1H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.50-7.30 (m, 10H), 3.98 (s, 1H), 1.48 (s, 3H); 13C NMR (100 MHz, CDCl3) *δ* 142.5, 136.1, 128.7, 128.4, 127.9, 127.7, 126.7, 125.4, 67.3, 63.3, 16.9.

**Table 2, entry 4:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> -17.3 (*c* 0.60, CHCl<sub>3</sub>) (83% ee); 1H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.28-7.23 (m, 1H), 7.14- 7.08 (m, 3H), 3.84 (dd,  $J = 4.2$ , 2.8 Hz, 1H), 3.14 (dd,  $J = 5.6$ , 4.2 Hz, 1H), 2.81 (dd,  $J = 5.6$ , 2.8 Hz, 1H), 2.35 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl3) *δ* 138.5, 137.7, 129.2, 128.6, 126.2, 122.9, 52.6, 51.4, 21.6.

**Table 2, entry 5:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> -13.8 (*c* 0.48, CHCl<sub>3</sub>) (80% ee); IR (NaCl) 1587, 1152 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) *<sup>δ</sup>* 7.30-7.24 (m, 1H), 7.01-6.92 (m, 3H), 5.18 (s, 2H), 3.84 (dd, *J* = 4.2, 2.8 Hz, 1H), 3.48 (s, 3H), 3.14 (dd, *J* = 5.6, 4.2 Hz, 1H),

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2.79 (dd,  $J = 5.6$ , 2.8 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 157.7, 139.6, 129.9, 119.2, 116.3, 113.4, 94.6, 56.3, 52.5, 51.4. Anal. Calcd for  $C_{10}H_{12}O_3$ : C, 66.65; H, 6.71. Found: C, 66.90; H, 6.94.

**Table 2, entry 6:** white solid;  $[\alpha]^{25}$ <sub>D</sub> +14.8 (*c* 1.80, CHCl<sub>3</sub>) (91% ee); 1H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.15-7.14 (m, 1H), 7.05- 7.02 (m, 1H), 6.70 (d,  $J = 8.4$  Hz, 1H), 3.86 (d,  $J = 4.2$  Hz, 1H), 3.48 (d, *J* = 4.2 Hz, 1H), 2.29 (s, 3H), 1.57 (s, 3H), 1.24 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) *δ* 150.5, 131.0, 130.6, 130.2, 119.9, 118.0, 73.0, 63.1, 51.3, 25.9, 22.7, 20.7.

**Table 2, entry 7:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> +106.9 (*c* 1.38, CHCl<sub>3</sub>) (96% ee); 1H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.34-7.31 (m, 2H), 7.13- 7.10 (m, 2H), 3.83 (s, 1H), 2.67-2.57 (m, 1H), 2.51-2.36 (m, 2H), 1.99-1.83 (m, 2H), 1.76-1.60 (m, 1H); 13C NMR (100 MHz, CDCl3) *δ* 135.5, 133.7, 128.6, 127.7, 66.9, 62.0, 31.5, 28.5, 12.6.

**Table 2, entry 8:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> +52.4 (*c* 0.67, CHCl<sub>3</sub>) (90% ee); IR (NaCl) 1604, 1497, 1455 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.38-7.32 (m, 2H), 7.30-7.24 (m, 3H), 4.02 (s, 1H),  $2.10-1.96$  (m, 1H),  $1.89-1.74$  (m, 3H),  $1.74-1.40$  (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl3) *δ* 137.4, 128.3, 127.7, 126.4, 72.8, 63.5, 34.2, 28.6, 25.5, 25.4. Anal. Calcd for C<sub>12</sub>H<sub>14</sub>O: C, 82.72; H, 8.10. Found: C, 82.56; H, 7.99.

**Table 2, entry 9:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub>  $-34.8$  (*c* 0.43, CHCl<sub>3</sub>) (88% ee); 1H NMR (400 MHz, CDCl3) *<sup>δ</sup>* 7.48-7.45 (m, 2H), 7.35- 7.30 (m, 3H), 3.65 (d,  $J = 4.4$  Hz, 1H), 3.27 (qd,  $J = 5.2$ , 4.4 Hz, 1H), 1.51 (d,  $J = 5.2$  Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 132.1, 129.0, 128.6, 122.4, 85.5, 84.3, 54.7, 46.2, 15.1.

**Table 2, entry 10:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> -31.2 (*c* 0.41, CHCl<sub>3</sub>) (90% ee); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.42 (dt,  $J = 4.4$ , 1.6 Hz, 1H), 3.13 (qd,  $J = 5.2$ , 4.4 Hz, 1H), 2.22 (td,  $J = 7.2$ , 1.6 Hz, 2H), 1.53 – 1.47 (m, 2H), 1.42 – 1.22 (m, 8H), 1.40 (d,  $J = 5.2$  Hz, 2H), 1.53-1.47 (m, 2H), 1.42-1.22 (m, 8H), 1.40 (d,  $J = 5.2$  Hz, 3H) 0.89 (t,  $J = 7.2$  Hz, 3H)<sup>, 13</sup>C NMR (100 MHz, CDCl<sub>2</sub>)  $\delta$ 3H), 0.89 (t,  $J = 7.2$  Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ <br>86.9. 75.1. 54.2. 46.1. 31.5. 28.7. 28.6. 22.8. 19.0. 14.9. 14.3. 86.9, 75.1, 54.2, 46.1, 31.5, 28.7, 28.6, 22.8, 19.0, 14.9, 14.3.

**Table 2, entry 11:** colorless oil;  $[\alpha]^{25}$ <sub>D</sub> -19.1 (*c* 0.33, CHCl<sub>3</sub>) (82% ee); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.02 (dt, *J* = 15.4, 4.4 Hz, 1H), 5.60 (ddt,  $J = 15.4$ , 7.6, 1.7 Hz, 1H), 4.22 (dd,  $J = 4.4$ , 1.7 Hz, 2H), 3.43 (dd,  $J = 7.6$ , 4.0 Hz, 1H), 3.22 (quint.,  $J = 5.4$ ) Hz, 1H), 1.29 (d,  $J = 5.4$  Hz, 3H), 0.91 (s, 9H), 0.07 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 136.3, 123.9, 63.2, 56.9, 54.7, 26.1,  $18.6, 13.6, -5.1.$ 

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**Supporting Information Available:** The data for the determination of the enantiomeric excess of the epoxides obtained with ketone **2c** along with the 1H and 13C NMR spectra of epoxides. This material is available free of charge via the Internet at http://pubs.acs.org.

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